

Review Article

Role of leptin and its receptors in the pathogenesis of thyroid cancer

Shahab Uddin, Azhar R Hussain, Abdul K Siraj, Omar S Khan, Prashant P Bavi, Khawla S Al-Kuraya

Human Cancer Genomic Research, Research Center, King Faisal Specialist Hospital and Research Center, Riyadh, Saudi Arabia

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Abstract: Leptin is a multifunctional adipose-derived cytokines that play a critical role in bodyweight homeostasis and energy balance. Recently, leptin and leptin receptor dysregulation have been reported in variety of malignant cells including thyroid. Leptin modulates growth and proliferation of cancer cells via activation of various growth and survival signaling pathways including JAK/STAT, PI3-kinase/AKT and/or Map kinases. In this review, current understanding of leptin's role in the pathogenesis of thyroid cancer has been described.

Keywords: Leptin, leptin receptors, signaling pathways, papillary thyroid carcinoma

Leptin and its receptors

Leptin, a product of the obese (*ob*) gene, is a 167-amino acid hormone [1, 2]. The *ob* gene is located on chromosome 7 [2]. Leptin acts through its receptor (Ob-R), which is encoded by the *db* gene [3]. Genetic deficiency of either leptin or Ob-R causes severe obesity and obesity-associated diabetes (Zhang et al 1994) [2]. Ob-R is a member of cytokine receptor family that including various interleukins, interferon- γ , erythropoietin, growth hormone and prolactin [3]. There are several leptin receptor variants (Ob-Ra through Ob-Rf) that are generated by alternative splicing. These variants share the same extracellular domain but have varied length of the transmembrane/cytoplasmic coding regions [4, 5]. The long Ob-Rb subtype (Ob-R_L) appears as the functional, signal-transducing isoform, responsible for the biological action of leptin (**Figure 1**). The biological function of the shorter Ob-R isoforms (Ob-R_S) remain to be characterized. Although leptin's precise sites of action are not known, its effect is thought largely mediated via hypothalamus. However, the wide expression of Ob-R_L suggests that leptin may also operate directly in other peripheral tissues [6]. There is now a significant amount of evidence implicating that leptin is active in the periphery [7].

Leptin mediated signaling

Leptin regulate multiple signaling pathways in various cancers. The major pathways that are regulated by leptin includes; JAK2/STAT3, erbB2, ERK, IRS and rho/rac pathways [8-13]. Binding of leptin with leptin receptors directly or indirectly activates these signaling pathways that involve kinase-induced phosphorylation of proteins. The leptin receptor is an external tyrosine kinase receptor; upon ligand binding each receptor can bind and activate the tyrosine kinase JAK2, which then cross-phosphorylates tyrosine residues in the other receptor in the dimer [14]. There is an absolute requirement of the intracellular cytokine box 1 motif of the receptor for activation of JAK2. This sequence is present in all the transmembrane isoforms. Most studies, however, have focused on signaling mediated by Ob-Rb, the only isoform which has conserved intracellular tyrosine residues and which is capable of activating the transcription factor STAT3 [15, 16]. In addition, only Ob-Rb has a cytokine box 2, which does not seem to be required for JAK2 activation, and a sequence of 15 amino acids downstream of box 1 that are required for optimal JAK2 activation [17, 18].

Leptin/Ob-R mediates its signaling via another

Leptin and its receptors in thyroid cancer

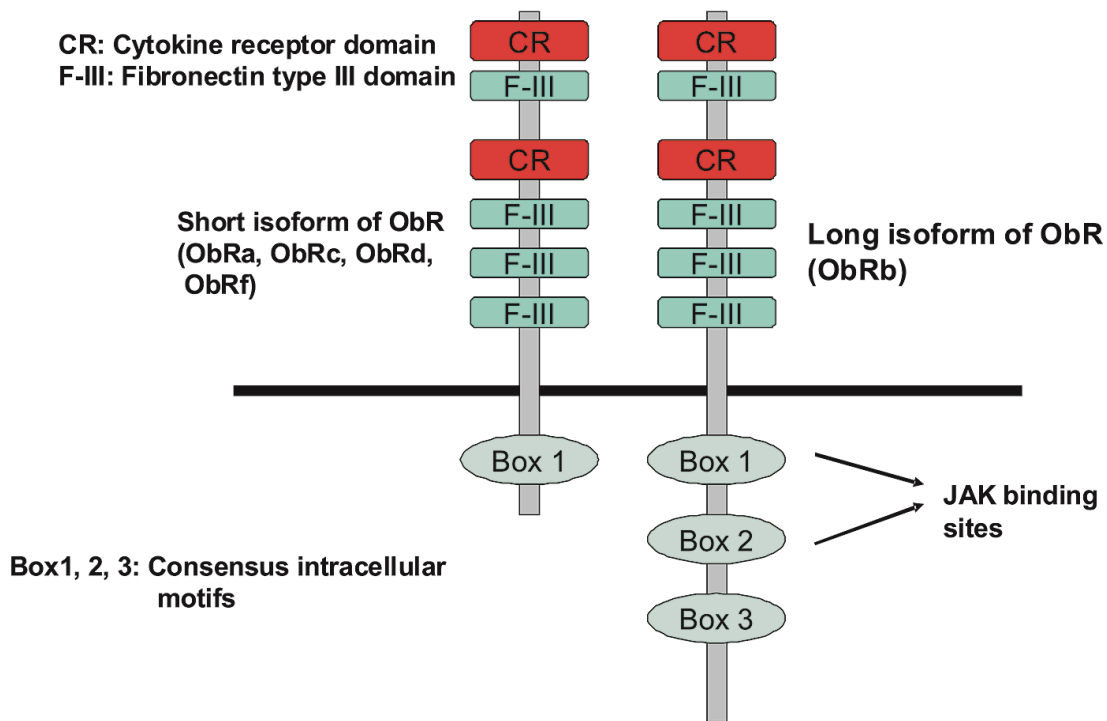


Figure 1. Schematic presentation of short and long form of leptin receptors.

major pathway, known as PI3K/AKT in oncogenesis in various tumor cells [19-23]. Leptin mediates Insulin receptor substrates (IRSs) phosphorylation by the intrinsic kinase activity of the receptor. Phosphorylation of IRSs increases the affinity by which they bind other signaling molecules, and initiates further steps on the pathway [24]. An important target of IRS molecules is phosphatidylinositol 3-kinase (PI 3-kinase) that generates inositoltrisphosphate (PIP3) [25]. IRSs exert PI 3-kinase activation through association with its regulatory subunit (p85), thus increasing the activity of the catalytic domain. Increased PIP3 levels lead to activation of PIP3-dependent serine/threonine kinases, such as PDK-1, 2, which can activate AKT, another serine/threonine kinase (**Figure 2**). AKT plays critical role in survival pathways by promoting glycolysis and maintaining a physiologic mitochondrial membrane potential [26]. In normal cells, growth factor and cytokines mediated signaling, recruits AKT through PH domain to the plasma membrane and activated by phosphorylation [27, 28]. Through its PH domain, AKT binds to PIP3, facilitating the activation of AKT by phosphoinositide-dependent kinase 1 and 2 (PDK1 and PDK2) by phosphorylation of threonine 308 and serine 473 [29].

Activated AKT has numerous targets that are important regulators of the cell cycle, the apoptotic pathway, and the translational and transcriptional machinery. AKT functions as an antiapoptotic factor through numerous mechanisms, including phosphorylation and inactivation of several proapoptotic factors such as Bad and caspase-9 [27]. Among these targets are the proapoptotic protein Bad, the cyclin-dependent kinase inhibitor p27^{kip1}, several forkhead family members, the mammalian target of rapamycin (mTOR), glycogen synthase kinase-3 β (GSK-3 β), and the I κ B kinases [30-34].

Recently we and others have shown that PTC cell lines expressed functional leptin receptors [35, 36]. Treatment of PTC cells with leptin enhances cell growth and prevented serum starved apoptosis [35]. Cheng SP et al [36] showed that leptin modulated the cell migration of thyroid cancer cell lines. These data suggests that oncogenic effect of leptin on PTC cells are due to a combination of cell proliferation and inhibition of apoptosis by leptin. In vitro experiments using PTC cell lines showed that leptin rapidly stimulates the PI3K pathway and induced the phosphorylation of AKT, thus activating of this key signal transduction pathway asso-

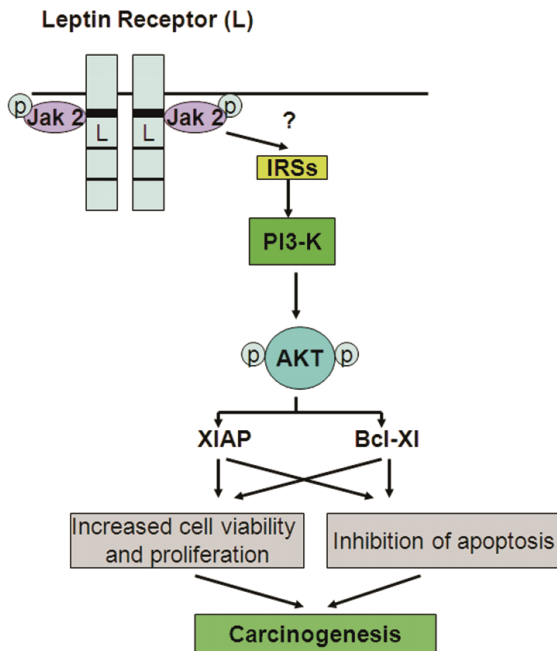


Figure 2: Schematic presentation showing leptin mediated signaling pathways. Upon leptin binding with extracellular domain of long isoform leptin receptor activates PI3-kinase/AKT pathway. The activated AKT in turn modulate the expression of antiapoptotic proteins XIAP and Bcl-xL leading to increased cell proliferation and inhibition of apoptosis resulting in initiation and progression of carcinogenesis.

ciated with cell growth. In addition, prevention of leptin-induced activation of PI3K with chemical inhibitors in turn significantly reduced the activation of AKT pathway. AKT provides a survival signal protecting cells from apoptosis induced by various stresses by multiple mechanisms such as phosphorylation of Bad, GSK3, FOXO transcription factors and caspase 9 [37, 38]. Phosphorylation of these proteins results in inactivation of their apoptotic functions. Leptin has been shown to activate AKT and its downstream targets such as GSK3, Foxo proteins in various tumor cells including prostate, colorectal and breast cancer cell lines [39-45]. Recently leptin has been shown to activate AKT and its downstream targets is a dose and time dependent manner in thyroid cancer cell lines BCPAP and TPC1 and inhibition of PI3K-kinase an upstream effector of AKT with LY294002 abolished leptin-mediated phosphorylation of AKT as well as cell proliferation. Thus we can hypothesize that leptin mediates its actions upstream of PI3K/AKT pathway. These data implicates that leptin mediated PTC carcinogenesis

occurs through deregulation of PI3K/AKT signaling pathway.

Leptin and leptin receptors in thyroid pathogenesis

The relationship between serum leptin and cancer risk has not been documented well in thyroid cancer. Recently, Akinci et al [46] using 43 PTC patients and 30 healthy control group reported that Serum leptin levels of TPC patients were significantly higher than in control group subjects. The leptin levels was decreased after total thyroidectomy compared to prethyroidectomy levels in most patients. These findings link leptin as a possible etiologic factor in thyroid carcinogenesis. However, the decreased post-thyroidectomy levels of leptin were still significantly higher than the control group levels of leptin, which may be related to hypothyroidism during the postoperative period for TSH stimulation. Leptin levels have been shown to increase with hypothyroidism [47]. Leptin is synthesised and secreted mainly by adipose cells, and its plasma levels in humans are strongly correlated with BMI [48]. However, Akinci et al (2009) study did not find significant difference in BMI in TPC and control groups. The leptin levels decreased postoperatively in all BMI subgroups in TPC patients.

Functional leptin receptors are expressed on diversified cancer cells derived from various solid tumors [49]. In these cancer cells, leptin able to mediates its growth and proliferative action via binding to its specific receptors [50]. Recently it has been shown that leptin receptors are expressed in PTC tumors. Using immunohistochemistry staining with Ob-R antibody on a large cohort of PTC tumor samples Ob-R protein expression was detected in 80% examined PTC [35]. Interestingly, Ob-R was significantly associated with older age, extra thyroid extension, larger tumor size, nodal metastasis, advanced stage and tall cell variant histologic subtype, thereby indicating that Ob-R overexpression identifies an aggressive phenotype of PTC. Furthermore patients with high Ob-R expressing tumors showed a significant poor disease free survival ($p < 0.0235$) as compared with reduced Ob-R expression. Raef H et al [51] have reported a fairly high number of patients with differentiated thyroid cancer (DTC) in Saudi Arabia had locally advanced disease at presentation and/or persistent disease after standard treatment. These findings speculate that high Ob-R

expression levels seen in around 80% of the Saudi Arabian PTCs could be one of the putative factors for this disease persistence and or recurrence. These findings therefore suggest that Ob-R expression is indeed associated aggressive phenotype and might be used as a marker for recurrent or persistent diseases of Middle Eastern PTC. Activated AKT (pAKT) protein expression was seen in 55% of PTC samples examined. However, no correlation was observed with Ob-R expression and AKT activation by IHC staining suggesting that the presence of other upstream signaling pathways can also be involved in the activation of AKT. It has been reported that leptin activate PI3-kinase/AKT pathway via forming as complex between insulin receptor substrates and JAK2 by SH2-B, a Jak2 interacting protein [52].

Leptin expression was found in PTC cell lines and depletion of Ob-R expression by using Ob-R small interference RNA abrogated leptin mediated activation of AKT and its downstream signaling in PTC cell lines. Ob-R overexpression has been found to be significantly associated with PIK3CA 110 alpha protein expression ($p < 0.0001$) in PTC tumors [35]. Leptin has been shown to mediate its action through the component of PI3-kinase/insulin signaling cascade [53]. XIAP is a member of anti apoptotic proteins, play a critical role in antiapoptotic function [54, 55]. XIAP is a physiological substrate of AKT. AKT interacts with XIAP and phosphorylates XIAP at serine 87. Phosphorylation of XIAP by AKT inhibits both its autoubiquitination and cisplatin-induced ubiquitination. These effects reduce XIAP degradation and the increased levels of XIAP are associated with decreased caspase 3 activity and programmed cell death [56]. Leptin receptor expression has been found to be significantly associated with XIAP expression in Saudi PTC tumors. In vitro studies small interference RNA of Ob-R in PTC cell lines downregulated XIAP transcript as well protein expression suggesting that leptin dysregulate PI3K/AKT signal transduction pathway leading to pathogenesis of PTC [35].

Conclusions

Evidence suggests that Leptin has multi effects in mediation of pathogenesis various malignant cells, including stimulation of tumor cell growth, migration and enhancement of angiogenesis. These actions of leptin play a role in tumor development and progression. In vitro studies

have shown that malignant cell lines including PTC cells express receptors for leptin and respond in a dose-dependent way to the administration of leptin. Elevated serum leptin levels seem to be associated with a higher risk for many cancers. There is some evidence that the increased risk for PTC. In various cancers, epidemiological reports show that higher levels of leptin may be related to poor prognosis than to increased risk for developing the disease. Leptin increases IGF and has growth potential in many in vitro studies and its effect seems permissive that it may exert its role only in the range of very low to normal leptin levels, with no additional effect in the range of normal to high leptin levels. Leptin modulates a number of growth and survival signaling pathways in other tumors such as prostate, Breast, ovarian, however, these pathways were not investigated in PTC. Therefore, further studies with more robust epidemiologic design, preferably prospective cohort investigations, are needed to evaluate in a more specific way, hypotheses generated by laboratory data.

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Conflicts of Interest

No conflicts of interest exist. The authors declare that they have no competing financial interest.

Address correspondence to: Shahab Uddin, PhD and Khawla S. Al-Kuraya, MD, FCAP Human Cancer Genomic Research, King Faisal Specialist Hospital and Research Center, MBC#98-16, P.O. Box 3354, Riyadh 11211, Saudi Arabia Tel: (966)-1-205-5167; Fax: (966)-1-205-5170; E-mail: Shahab@kfshrc.edu.sa; E-mail: kkuraya@kfshrc.edu.sa

References

- [1] Baratta M. Leptin-from a signal of adiposity to a hormonal mediator in peripheral tissues. *Med Sci Monit* 2002; 8: RA282-RA292.
- [2] Zhang Y, Proenca R, Maffei M, Barone M, Leopold L and Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature* 1994; 372: 425-432.
- [3] Tartaglia LA, Dembski M, Weng X, Deng N, Culpepper J, Devos R, Richards GJ, Campfield

- LA, Clark FT, Deeds J, Muir C, Sanker S, Moriarty A, Moore KJ, Smutko JS, Mays GG, Wool EA, Monroe CA and Tepper RI. Identification and expression cloning of a leptin receptor, OB-R. *Cell* 1995; 83: 1263-1271.
- [4] Chua SC Jr, Chung WK, Wu-Peng XS, Zhang Y, Liu SM, Tartaglia L and Leibel RL. Phenotypes of mouse diabetes and rat fatty due to mutations in the OB (leptin) receptor. *Science* 1996; 16: 994-996.
- [5] Ahima RS and Flier JS. Leptin. *Annu Rev Physiol* 2000; 62: 413-437.
- [6] Lin J, Barb CR, Matteri RL, Kraeling RR, Chen X, Meinersmann RJ and Rampacek GB. Long form leptin receptor mRNA expression in the brain, pituitary, and other tissues in the pig. *Domest Anim Endocrinol* 2000; 19: 53-61.
- [7] Caprio M, Fabbrini E, Isidori AM, Aversa A and Fabbri A. Leptin in reproduction. *Trends Endocrinol Metab* 2001; 12: 65-72.
- [8] Ghilardi N, Ziegler S, Wiestner A, Stoffel R, Heim MH and Skoda RC. Defective STAT signaling by the leptin receptor in diabetic mice. *Proc Natl Acad* 1996; 93: 6231-6235.
- [9] Myers MG Jr. Leptin receptor signaling and the regulation of mammalian physiology, *Rec Prog Horm Res* 2004; 59: 287-304.
- [10] Eisenberg A, Biener E, Charlier M, Krishnan RV, Djiane J, Herman B and Gertler B. Transactivation of erbB2 by short and long isoforms of leptin receptors. *FEBS Lett* 2004; 565: 139-142.
- [11] Bjorbaek C, Uotani S, da Silva, B and Flier JS. Divergent signaling capacities of the long and short isoforms of the leptin receptor. *J Biol Chem* 1997; 272: 32686-32695.
- [12] Sobhani I, Bado A, Vissuzaine C, Buyse M, Kermorgant S, Laigneau J, Attoub PS, Lehy T, Henin D, Mignon M and Lewin MJ. Leptin secretion and leptin receptor in the human stomach. *Gut* 2000; 47: 178-183.
- [13] Bahrenberg G, Behrmann I, Barthel A, Hekerman P, Heinrich PC, Joost HG and Becker W. Identification of the critical sequence elements in the cytoplasmic domain of leptin receptor isoforms required for Janus kinase/signal transducer and activator of transcription activation by receptor heterodimers. *Mol Endocrinol* 2002; 16: 859-872.
- [14] Jaffe T and Schwartz B. Leptin promotes motility and invasiveness in human colon cancer cells by activating multiple signal-transduction pathways. *Int J Cancer* 2008; 123: 2543-2556.
- [15] Jiang L, Li Z and Rui L. Leptin stimulates both JAK2-dependent and JAK2-independent signaling pathways. *J Biol Chem* 2008; 283: 28066-28073.
- [16] Banks AS, Davis SM, Bates SH and Myers MG Jr. Activation of downstream signals by the long form of the leptin receptor. *J Biol Chem* 2000; 275: 14563-14572.
- [17] Bates SH, Stearns WH, Dundon TA, Schubert M, Tso AW, Wang Y, Banks AS, Lavery HJ, Haq AK, Maratos-Flier E, Neel BG, Schwartz MW and Myers MG Jr. STAT3 signalling is required for leptin regulation of energy balance but not reproduction. *Nature* 2003; 421: 856-859.
- [18] da Silva BA, Bjorbaek C, Uotani S and Flier JS. Functional properties of leptin receptor isoforms containing the gln->pro extracellular domain mutation of the fatty rat. *Endocrinology* 1998; 39: 3681-3690.
- [19] Bjorbaek C, Uotani S, da Silva B and Flier JS. Divergent signaling capacities of the long and short isoforms of the leptin receptor. *J Biol Chem* 1997; 272: 32686-32695.
- [20] Hoda MR, Keely SJ, Bertelsen LS, Junger WG, Dharmasena D and Barrett KE. Leptin acts as a mitogenic and antiapoptotic factor for colonic cancer cells. *Br J Surg* 2007; 94: 346-354.
- [21] Ogunwobi OO and Beales IL. The anti-apoptotic and growth stimulatory actions of leptin in human colon cancer cells involves activation of JNK mitogen activated protein kinase, JAK2 and PI3 kinase/AKT. *Int J Colorectal Dis* 2007; 22: 401-409.
- [22] Saxena NK, Sharma D, Ding X, Lin S, Marra F, Merlin D and Anania FA. Concomitant activation of the JAK/STAT, PI3K/AKT, and ERK signaling is involved in leptin-mediated promotion of invasion and migration of hepatocellular carcinoma cells. *Cancer Res* 2007; 67: 2497-2507.
- [23] Sharma D, Saxena NK, Vertino PM and Anania FA. Leptin promotes the proliferative response and invasiveness in human endometrial cancer cells by activating multiple signal-transduction pathways. *Endocr Relat Cancer* 2006; 13: 629-640.
- [24] Rordorf-Nikolic T, Van Horn DJ, Chen D, White MF, Backer JM. Regulation of phosphatidylinositol 3'-kinase by tyrosyl phosphoproteins. Full activation requires occupancy of both SH2 domains in the 85-kDa regulatory subunit. *J Biol Chem* 1995; 270: 3662-3666.
- [25] Kane LP and Weiss A. The PI-3 kinase/AKT pathway and T cell activation: pleiotropic pathways downstream of PIP3. *Immunol Rev* 2003; 192: 7-20.
- [26] Plas DR, Talapatra S, Edinger AL, Rathmell JC and Thompson CB. AKT and Bcl-xL promote growth factor-independent survival through distinct effects on mitochondrial physiology. *J Biol Chem* 2001; 276: 12041-12048.
- [27] Franke TF, Hornik CP, Segev L, Shostak GA and Sugimoto C. PI3K/AKT and apoptosis: size matters. *Oncogene* 2003; 22: 8983-8998.
- [28] Matsuzawa A and Ichijo H. Stress-responsive protein kinases in redox-regulated apoptosis signaling. *Antioxid Redox Signal* 2005; 7: 472-481.
- [29] Anderson KE and Jackson SP. Class I phos-

- phoinositide 3-kinases. *Int J Biochem Cell Biol* 2003; 35: 1028-1033.
- [30] del Peso L, Gonzalez-Garcia M, Page C, Herrera R and Nunez G. Interleukin-3-induced phosphorylation of BAD through the protein kinase AKT. *Science* 1997; 278: 687-689.
- [31] Gajewski TF and Thompson CB. Apoptosis meets signal transduction: elimination of a BAD influence. *Cell* 1996; 87: 589-592.
- [32] Liang J, Zubovitz J, Petrocelli T, Kotchetkov R, Connor MK, Han K, Lee JH, Ciarallo S, Catzavelos C, Beniston R, Franssen E and Slingerland JM. PKB/AKT phosphorylates p27, impairs nuclear import of p27 and opposes p27-mediated G1 arrest. *Nat Med* 2002; 8: 1153-1160.
- [33] Cross DA, Alessi DR, Cohen P, Andjelkovich M and Hemmings BA. Inhibition of glycogen synthase kinase-3 by insulin mediated by protein kinase B. *Nature* 1995; 378: 785-789.
- [34] Hay N. The AKT-mTOR tango and its relevance to cancer. *Cancer Cell* 2005; 8: 179-183.
- [35] Uddin S, Bavi P, Siraj AK, Ahmed M, Al-Rasheed M, Hussain AR, Ahmed M, Amin T, Alzahrani A, Al-Dayel F, Abubaker J, Bu R and Al-Kuraya KS. Leptin-R and its association with PI3K/AKT signaling pathway in papillary thyroid carcinoma. *Endocr Relat Cancer* 2010; 17: 191-202.
- [36] Cheng SP, Yin PH, Chang YC, Lee CH, Huang SY and Chi CW. Differential roles of leptin in regulating cell migration in thyroid cancer cells. *Oncol Rep* 2010; 23: 1721-1727.
- [37] Brunet A, Datta SR and Greenberg ME. Transcription-dependent and -independent control of neuronal survival by the PI3K-AKT signaling pathway. *Curr Opin Neurobiol* 2001; 11: 297-305.
- [38] Datta R, Oki E, Endo K, Biedermann V, Ren J and Kufe D. XIAP regulates DNA damage-induced apoptosis downstream of caspase-9 cleavage. *J Biol Chem* 2000; 275: 31733-31738.
- [39] Huang CY, Yu HS, Lai TY, Yeh YL, Su CC, Hsu HH, Tsai FJ, Tsai CH, Wu HC and Tang CH. Leptin increases motility and integrin up-regulation in human prostate cancer cells. *J Cell Physiol* 2011; 226: 1274-1282.
- [40] Uddin S, Bavi PP, Hussain AR, Alsbeih G, Al-Sanea N, Abduljabbar A, Ashari LH, Alhounoud S, Al-Dayel F, Ahmed M and Al-Kuraya KS. Leptin receptor expression in Middle Eastern colorectal cancer and its potential clinical implication. *Carcinogenesis* 2009; 30: 1832-1840.
- [41] Paik SS, Jang SM, Jang KS, Lee KH, Choi D and Jang SJ. Leptin expression correlates with favorable clinicopathologic phenotype and better prognosis in colorectal adenocarcinoma. *Ann Surg Oncol* 2009; 16: 297-303.
- [42] Uddin S, Bu R, Ahmed M, Abubaker J, Al-Dayel F, Bavi P and Al-Kuraya KS. Overexpression of leptin receptor predicts an unfavorable outcome in Middle Eastern ovarian cancer. *Mol Cancer* 2009; 8: 74.
- [43] Choi JH, Park SH, Leung PC and Choi KC. Expression of leptin receptors and potential effects of leptin on the cell growth and activation of mitogen-activated protein kinases in ovarian cancer cells. *J Clin Endocrinol Metab* 2005; 90: 207-210.
- [44] Uddin S, Bu R, Ahmed M, Hussain AR, Ajarim D, Al-Dayel F, Bavi P and Al-kuraya KS. Leptin receptor expression and its association with PI3K/AKT signaling pathway in diffuse large B-cell lymphoma. *Leuk Lymphoma* 2010; 51: 1305-1314.
- [45] Frankenberry KA, Skinner H, Somasundar P, McFadden DW and Vona-Davis LC. Leptin receptor expression and cell signaling in breast cancer. *Int J Oncol* 2006; 28: 985-993.
- [46] Akinci M, Kosova F, Cetin B, Aslan S, Ari Z and Cetin A. Leptin levels in thyroid cancer. *Asian J Surg* 2009; 32: 216-223.
- [47] Botella-Carretero JI, Alvarez-Blasco F, Sancho J and Escobar-Morreale HF. Effects of thyroid hormones on serum levels of adipokines as studied in patients with differentiated thyroid carcinoma during thyroxine withdrawal. *Thyroid* 2006; 16: 397-402.
- [48] Considine RV, Sinha MK, Heiman ML, Kriaucinas A, Stephens TW, Nyce MR, Ohannesian JP, Marco CC, McKee LJ, Bauer TL and Caro JF. Serum immunoreactive-leptin concentrations in normal-weight and obese humans. *N Engl J Med* 1996; 334: 292-295.
- [49] Cheng SP, Chi CW, Tzen CY, Yang TL, Lee JJ, Liu TP and Liu CL. Clinicopathologic significance of leptin and leptin receptor expressions in papillary thyroid carcinoma. *Surgery* 2010; 147: 847-853.
- [50] Mantzos F, Vanakara P, Samara S, Wosniak G, Kollia P, Messinis I and Hatzitheofilou C. Leptin receptor expression in neoplastic and normal ovarian and endometrial tissue. *Eur J Gynaecol Oncol* 2011; 32: 84-86.
- [51] Raef H, Alfadhli E, Al-Hajjaj A, Malabu UH, Al-Sobhi S, Rifai A and Al Nuaim A. High rate of persistent/recurrent disease among patients with differentiated thyroid cancer in Saudi Arabia: factors affecting nonremission. *Ann Saudi Med* 2008; 28: 277-281.
- [52] Duan C, Li M and Rui L. SH2-B promotes insulin receptor substrate 1 (IRS1)-and IRS2-mediated activation of the phosphatidylinositol 3-kinase pathway in response to leptin. *J Biol Chem* 2004; 279: 43684-43691.
- [53] Huang CY, Yu HS, Lai TY, Yeh YL, Su CC, Hsu HH, Tsai FJ, Tsai CH, Wu HC and Tang CH. Leptin increases motility and integrin up-regulation in human prostate cancer cells. *J Cell Physiol* 2011; 226: 1274-1282.
- [54] Dan HC, Sun M, Kaneko S, Feldman RI, Nicolsia SV, Wang HG, Tsang BK and Cheng JQ.

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- AKT phosphorylation and stabilization of X-linked inhibitor of apoptosis protein (XIAP). *J Biol Chem* 2004; 279: 5405-5412.
- [55] Uddin S, Hussain AR, Siraj AK, Manogaran PS, Al-Jomah NA, Moorji A, Atizado V, Al-Dayel F, Belgaumi A, El-Solh H, Ezzat A, Bavi P and Al-Kuraya KS. Role of phosphatidylinositol 3'-kinase/AKT pathway in diffuse large B-cell lymphoma survival. *Blood* 2006; 108: 4178-4186.
- [56] Asselin E, Mills GB and Tsang BK. XIAP regulates AKT activity and caspase-3-dependent cleavage during cisplatin-induced apoptosis in human ovarian epithelial cancer cells. *Cancer Res* 2001; 61: 1862-1868.