

## Case Report

# Co-existence of t(9;22) and t(8;21) in primary blast phase of chronic myelogenous leukemia: clinical experience and literature review

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**Abstract:** Rare chronic myelogenous leukemia (CML) patients manifested as the primary blast phase without a chronic and accelerated phase. The occurrence of a t(8;21) translocation in secondary blast phase of CML or Philadelphia chromosome positive acute myelogenous leukemia (Ph+ AML) has been reported previously. No case of primary blast phase of chronic myelogenous leukemia (CML-BP) bearing one clone with t(9;22) and t(8;21) simultaneously has been reported. One Chinese patient presenting with extensive spontaneous ecchymosis and enlarged spleen diagnosed as acute myelogenous leukemia (AML) by smear and immunophenotype was given chemotherapy including daunorubicin 3 days and cytarabine 7 days without a tyrosine kinase inhibitor (TKI) drug at the beginning. Fresh frozen plasma and 4-factor prothrombin complex concentrate was also transfused for coagulation disorder. However, fusion genes BCR/ABL p210 and AML1/ETO were both positive and karyotype analysis showed the abnormalities of t(9;22) and t(8;21) in the same clones. Bone marrow aspirate on 7th day of chemotherapy indicated hypocellularity with 45% blasts remaining. Cytarabine was prolonged to nine days combined with imatinib 600 mg per day. His bone marrow aspirate after complete remission revealed t(8;21) clones disappearing, especially FISH of bone marrow smear detecting the BCR/ABL fusion signals in the basophilic erythroblasts, which confirmed his diagnosis as primary blast phase of CML rather than Ph+ AML. Thus, we report for the first time one patient diagnosed as primary blast phase of CML presenting with t(9;22) and t(8;21) simultaneously.

**Keywords:** Chronic myelogenous leukemia, primary blast phase, coagulation disorder, case report, chromosome translocation

## Introduction

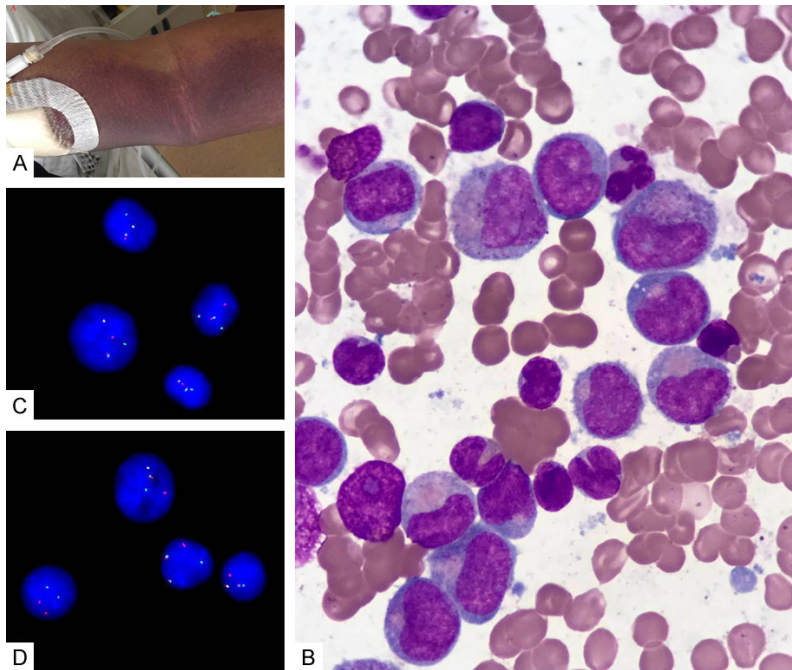
CML is a clonal myeloproliferative disorder of pluripotent hematopoietic stem cells characterized by specific hematologic and chromosomal changes [1]. The clinical course of patients with CML generally is divided into three phases: chronic phase, accelerated phase, and blast phase. The specific chromosomal abnormality is the Philadelphia chromosome (Ph), which results from a translocation involving the *abl* gene at chromosome 9q34 and the *bcr* gene at chromosome 22q11 [2]. The encoded chimeric protein, BCR/ABL is the target of TKI drug therapy [3]. Rare CML patients manifested as primary blast phase without a chronic and accel-

erated phase [4]. The t(8;21)(q22;q22) typically is associated with a distinct type of AML with characteristic morphologic features and a favorable clinical outcome [5]. The occurrence of this translocation in secondary blast phase of CML or Ph+ AML has been reported previously [6-12]. Here we report for the first time one patient diagnosed in the primary blast phase of chronic myelogenous leukemia bearing one clone with t(9;22) and t(8;21) simultaneously.

## Case presentation

A 45-year-old Chinese man with chief complaints of jaw pain for two months was admitted into our hospital on June 6, 2017. No positive

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**Figure 1.** Clinical manifestations, morphology and FISH results before treatment. A. Obvious bleeding of the arm. B. Bone marrow blasts showing abundant granular cytoplasm with clear nucleoli and perinuclear clearing. C. In cells with t(8;21), the hybridization produces fused yellow signals (AML1/ETO), orange signals (ETO), and green signals (AML1). D. In cells with t(9;22), the hybridization produces fused yellow signals (BCR/ABL), red signals (native ABL), and 1 green signal (native BCR).

past history was obtained. One week before admission extensive spontaneous ecchymosis and subcutaneous nodules were found. Enlarged spleen was 5 cm under the costal margin. Complete blood count showed hemoglobin 107 g/L, thrombocytopenia  $49 \times 10^9/L$ , leukocytosis  $172.26 \times 10^9/L$  with 37% neutrophils, 2% monocytes, 9% lymphocytes, 3% myelocytes, 4% metamyelocytes, 45% blasts, and coagulation testing showed partial thromboplastin time 20.2 sec (11-15), activated partial thromboplastin time 28.5 sec (20-40), fibrinogen 3.75 g/L, thrombin time 20.4 sec (14-21), d-dimer 28 mg/L (**Figure 1A**). Bone marrow smear disclosed markedly increased cellularity with 33% myeloblasts, 18% promyelocytes, 18% myelocytes, 6% metamyelocytes, 20% mature neutrophils, 1% eosinophils, 0.5% basophils, 8% lymphocytes, and 0.5% normoblasts (**Figure 1B**). Flow cytometric immunophenotyping of bone marrow showed the blasts were positive for CD33, CD123, CD38, MPO, partially positive for CD15, CD13, CD11b, CD9 and were negative for CD34, HLA-DR, CD19. He was given a regimen including daunorubicin 80 mg/

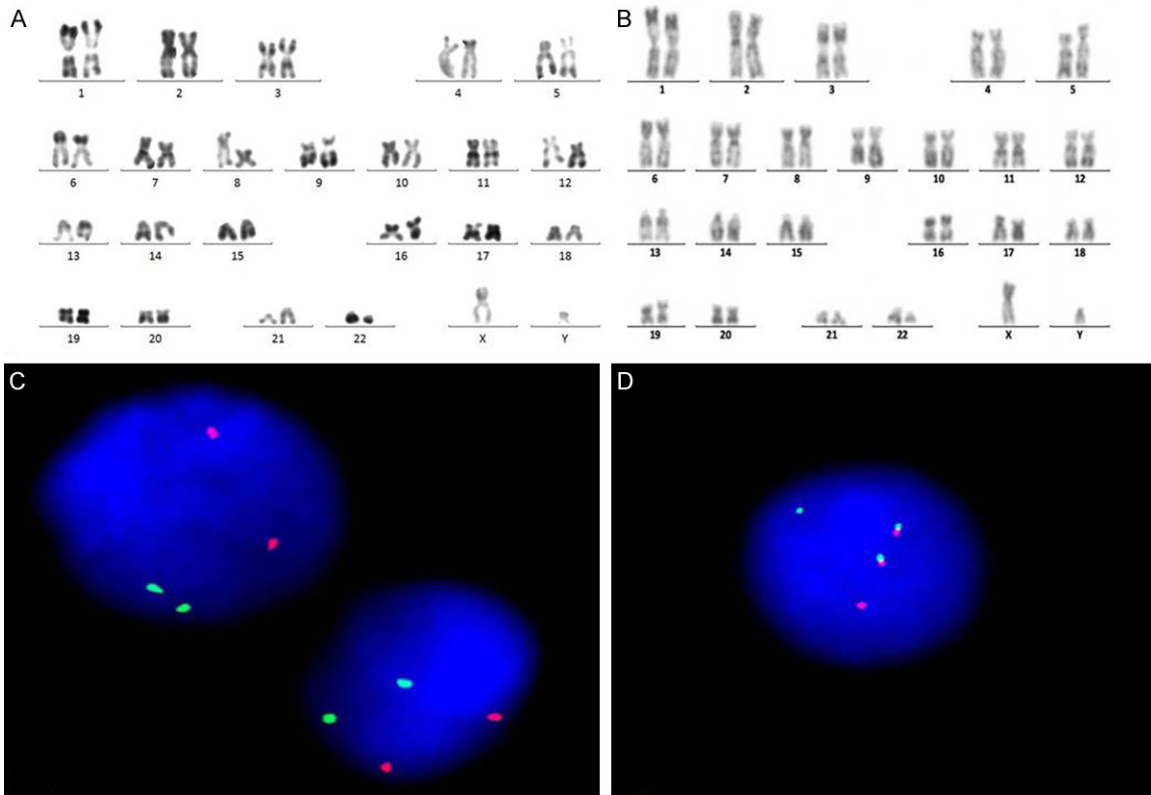
$m^2 \times 3$  day and cytarabine  $100 \text{ mg}/m^2 \times 7$  day on hospital day 3 after admission. At the beginning, fresh frozen plasma and 4-factor prothrombin complex concentrate was transfused for coagulation disorder. Reverse transcription polymerase chain reaction (RT-PCR) of AML fusion genes BCR/ABL p210 and AML1/ETO were both positive. The BCR/ABL and ABL1/ETO nuclear fusion signals were also detected by *in situ* hybridization (FISH) in the interphase cells (**Figure 1C, 1D**). Karyotype analysis revealed chromosomal abnormalities t(9;22) and the t(8;21) in all 15 evaluated mitoses (**Figure 2A**). His T3-15l mutation was negative. Bone marrow aspirate on 7th day of chemotherapy indicated hypocellularity with 45% blasts remaining. Then cytarabine was prolonged to nine days com-

combined with imatinib 600 mg/day. The third bone marrow aspirate on July 7, 2017 showed hypocellularity without excess blasts and fusion gene BCR/ABL was still positive but AML1/ETO turned out as negative, which indicated his disease was back to the chronic phase of CML. After complete remission, the karyotype analysis of fourth aspirate showed t(9;22) abnormality without t(8;21) (**Figure 2B**), correlating with FISH results (**Figure 2C, 2D**). BCR-ABL fusion signal was also detected in the basophilic erythroblasts on the bone marrow smear which definitely confirmed his diagnosis of CML-BP, not Ph+ AML (**Figure 3A, 3B**). Because of donor and financial limitation, he could not receive allogeneic stem cell transplantation and died from early relapse and resistance to chemotherapy two months later.

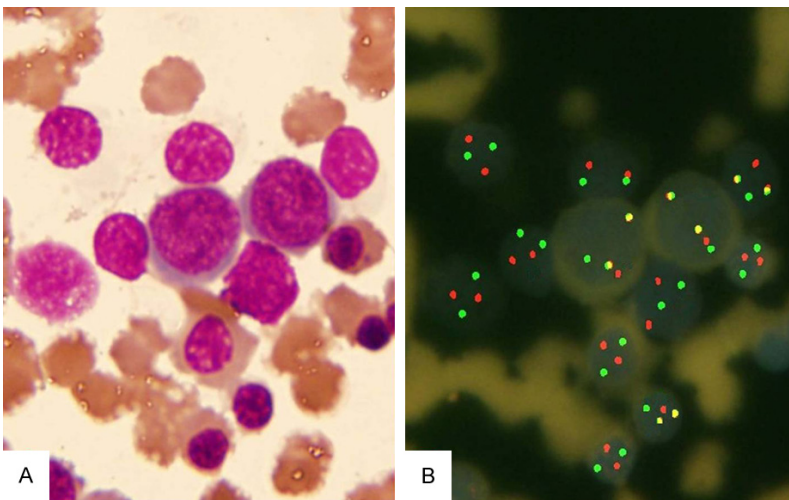
### Discussion

As to our literature review, this is the first case diagnosed as primary blast phase of CML with co-existence of t(9;22) and t(8;21). Early in 1978, Dr. Francesconi found a 13-year old patient diagnosed as AML with t(8;21); howev-

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**Figure 2.** Karyotype of bone marrow before and after chemotherapy and FISH results of bone marrow after chemotherapy. A. Karyotype of the bone marrow before treatment showing t(9;22) and t(8;21) abnormality in the same leukemic clone cells. B. Karyotype of the bone marrow after treatment: t(9;22) without t(8;21) abnormality. C. FISH showing disappearance of fused yellow signals (AML1/ETO) after effective treatment. D. FISH showing remaining fused yellow signals (BCR/ABL) after effective treatment.



**Figure 3.** FISH results of the BCR/ABL signals in the basophilic erythroblasts on the bone marrow smear. A. Two basophilic erythroblasts on the bone marrow smear. B. FISH of bone marrow smear showing fused yellow signals (BCR/ABL) in the two basophilic erythroblasts.

py were given, chromosome analysis showed presence of a Ph<sup>+</sup> chromosome [6]. Xue et al [7] reported one case of basophilic leukemia bearing simultaneous translocations of t(8;21) and t(9;22); however, differential diagnosis between CML and Ph<sup>+</sup> AML was undetermined. The presence of a t(8;21) alteration can be detected as an additional chromosome in blast phase of CML with disease progression [8, 9]. Ammatuna et al have described cases of acute myeloid leukemia with simultaneous occurrence of t(9;22) and t(8;21) [10-12].

er, treatment was unsatisfactory and eight months after diagnosis and courses of thera-

It is difficult to distinguish primary blast phase of CML from Ph<sup>+</sup> AML; however, history of ante-



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cedent blood anomaly, basophilia >2% in white blood cells, and 100% BCR/ABL metaphases would be helpful for diagnosis of CML [4]. CML originates from pluripotent hematopoietic stem cells and BCR-ABL is the main driving event in CML, sufficient to initiate the disease while BCR-ABL in AML is a secondary event [11, 13]. The presence of the Ph chromosome in erythroblasts, neutrophils, eosinophils, basophilic granulocytes, macrophages and megakaryocytes is specific for CML [14]. For this reason, when a CML-BP patient is back to chronic phase, Ph chromosome is always still present. In our case, the Ph<sup>+</sup> clone remained while disappearance of t(8;21) clones after effective treatment indicated a CML-BP diagnosis. Identification of BCR/ABL fusion signals in these cells in the bone marrow or peripheral blood smears by FISH is a well way to differentiate CML from AML [15, 16], just as the method we used in our case.

Patients with CML in primary blast crisis should be treated with a TKI drug with or without chemotherapy, with the goal of reversion to chronic phase and proceeding to allogeneic stem cell transplantation as quickly as possible [17]. Our patient was treated according to acute myelogenous leukemia by his bone marrow morphology and immunophenotype at the beginning. However, coagulation disorder deteriorated during chemotherapy. More importantly, the second bone aspirate showed 45% blasts remaining, but he achieved complete remission after adding a TKI drug, which indicated the importance of TKI drugs in treating CML-BP.

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Written informed consent was obtained from the patient for publication of this case report.

### Disclosure of conflict of interest

None.

### Abbreviations

CML, Chronic myelogenous leukemia; Ph<sup>+</sup> AML, Philadelphia chromosome positive Acute Mye-

logenous Leukemia; AML, Acute Myelogenous Leukemia; TKI, Tyrosine kinase inhibitor; CML-BP, blast phase of chronic myelogenous leukemia; Ph, Philadelphia chromosome; RT-PCR, reverse transcription polymerase chain reaction; FISH, fluorescence in situ hybridization.

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